Appendix A. Supplementary data

Broadband Laser-based Mid-Infrared Spectroscopy employing a Quantum Cascade Detector for Milk Protein Analysis

Alicja Dabrowska,[†] Mauro David,[‡] Stephan Freitag,[†] Aaron Maxwell Andrews,[‡] Gottfried Strasser,[‡] Borislav Hinkov,[‡] Andreas Schwaighofer,^{†,*} and Bernhard Lendl^{†,*}

† Institute of Chemical Technologies and Analytics, Technische Universität Wien, Getreidemarkt 9/164-UPA, 1060 Vienna, Austria

‡ Institute of Solid State Electronics & Center for Micro- and Nanostructures, Technische Universität Wien, Gußhausstrasse 25-25a, 1040 Vienna, Austria

DESCRIPTION OF CONTENTS:

Figure S1. Signal and noise levels of the EC-QCL – QCD setup at different transmission path lengths.

- Figure S2. Spectral FTIR characterization of a ridge QCD at room temperature.
- Figure S3. Calibration curves obtained for bovine milk proteins by the EC-QCL-QCD setup.

Equation S1. The degree of spectral overlap.

Figure S4. Absorption spectra of ternary milk proteins recorded with the EC-QCL-QCD setup for multivariate quantitation.

Table S1. Protein concentrations in calibration samples and figures of merit obtained by PLS.

Figure S5. Nominal and predicted milk protein concentration values obtained by PLS.



Figure S1. Signal and noise levels of the EC-QCL – QCD setup at different transmission path lengths. Signal was defined as the absorbance at the 1632 cm⁻¹ (maximum of the amide I band for β -lactoglobulin) and the RMS noise level of the 100% lines of water between 1700 and 1600 cm⁻¹. The measurements were performed at acquisition times of ~45 s (300 scans) and spectral resolution of 2.6 cm⁻¹.



Figure S2. Spectral FTIR characterization of the ridge QCD at room temperature. Gray-shaded area corresponds to the EC-QCL tunability region. Gray squares correspond to the responsivity of the QCD measured by an EC-QCL at different wavenumbers.



Figure S3. Calibration curves obtained for bovine milk proteins by the EC-QCL-QCD setup. Quantitative analysis was performed by evaluation of the height of the amide I band maxima at 1632, 1654 and 1651 cm⁻¹ for β -lactoglobulin, α -lactalbumin and casein, respectively. Five measurements were averaged per concentration.

To quantitatively compare the agreement between the spectral components, the degree of spectral overlap (s_{12}) is calculated:

$$s_{12} = \frac{\|s_1^T s_2\|}{\|s_1\| \|s_2\|'}$$
(S1)

where s_1 is the spectrum obtained by EC-QCL-QCD setup and s_2 is the spectrum measured by an FTIR instrument. The value of s_{12} ranges from 0 to 1, where 0 indicates lack of overlap and 1 suggests complete spectral overlap [1].

[1] M.J. Culzoni, H.C. Goicoechea, G.A. Ibañez, V.A. Lozano, N.R. Marsili, A.C. Olivieri, A.P. Pagani, Second-order advantage from kineticspectroscopic data matrices in the presence of extreme spectral overlapping. A multivariate curve resolution-Alternating least-squares approach, Anal. Chim. Acta. 614 (2008) 46–57. https://doi.org/10.1016/j.aca.2008.03.013.



Figure S4. Absorption spectra of ternary milk proteins recorded with the EC-QCL-QCD setup for multivariate quantitation.

Calibration sample	β -LG (mg mL ⁻¹)		α -LA (mg mL ⁻¹)		Cas (mg mL ⁻¹)		Total protein (mg mL ⁻¹)	
	Nominal	Predicted	Nominal	Predicted	Nominal	Predicted	Nominal	Predicted
C-01	1.50	1.49	3.00	3.30	1.00	1.37	5.50	5.57
C-02	7.00	6.75	1.00	0.55	1.50	1.97	9.50	9.41
C-03	3.00	2.73	4.50	4.04	2.00	1.92	9.50	8.55
C-04	5.50	5.79	8.00	8.15	2.50	2.71	16.00	15.67
C-05	2.50	2.61	2.50	2.66	3.00	2.73	8.00	8.17
C-06	8.00	7.58	6.00	5.73	3.50	2.87	17.50	16.09
C-07	2.00	1.98	5.50	5.95	4.00	3.81	11.50	11.48
C-08	7.50	7.85	9.00	9.06	4.50	5.07	21.01	20.67
C-09	3.50	3.28	10.00	9.87	5.00	5.29	18.51	17.82
C-10	6.00	6.76	2.00	2.51	5.50	6.19	13.50	14.60
C-11	1.00	1.31	4.00	3.95	6.00	6.07	11.00	12.53

Table S1. Protein concentrations in calibration samples and figures of merit obtained by PLS.





Figure S5. Nominal and predicted milk protein concentration values obtained by PLS.