

# Biomarker detection: from total serum to tissue specific N-glycan analysis

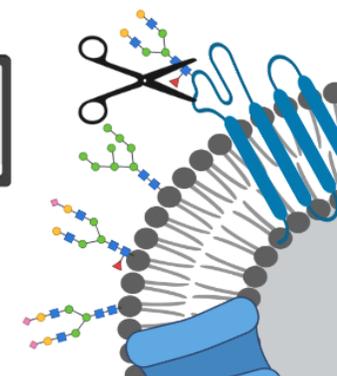
Davide Ret<sup>a,b\*</sup>, Alessio Gentile<sup>a,b</sup>, Daniel García de Otazo Hernández<sup>a,b</sup>, Lisa Ying<sup>a</sup>, Veronica Montia<sup>a</sup>, Haidi Jakovic, Philipp Gritsch<sup>a</sup>, Arvand Haschemi<sup>c</sup>, Eva Untersmayr<sup>b</sup>

<sup>a</sup> *Institute of Applied Synthetic Chemistry, TU Wien, 1060 Vienna, Austria,*

<sup>b</sup> *Institute of Pathophysiology and Allergy Research, Medical University of Vienna, 1090 Vienna, Austria*

<sup>c</sup> *Department of Laboratory, Medicine Medical University of Vienna, 1090 Vienna, Austria*

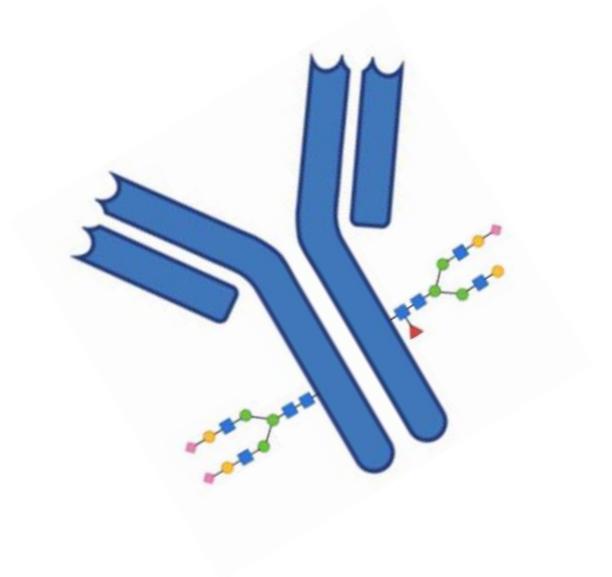
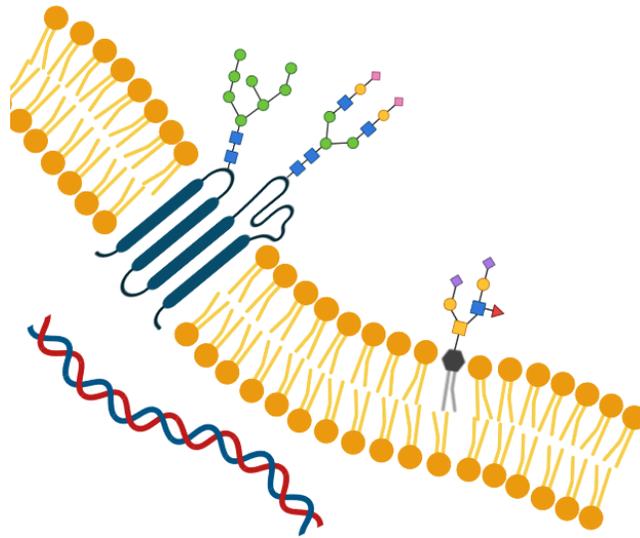
davide.ret@tuwien.ac.at



# Glycans

key components in most **biological** and **pathological** processes:

- antigen recognition
- inflammation
- cytotoxicity
- cancer
- viral and bacterial infections.



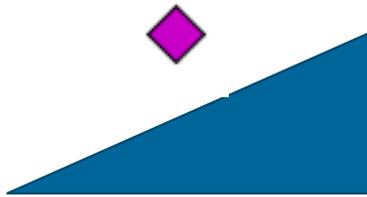
# glycosylation:

Biological age



glycan clock

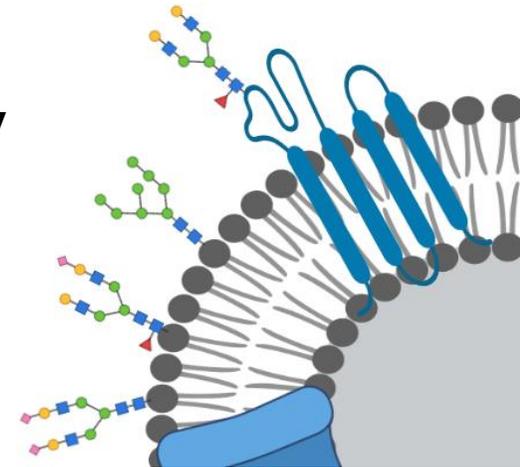
inflammatory  
regulators



receptors  
(glycan-lectins)



species and  
tissue specificity

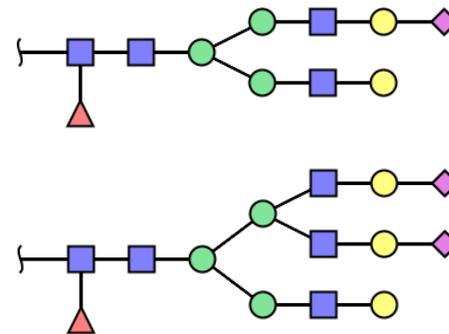


# types of glycosylation:

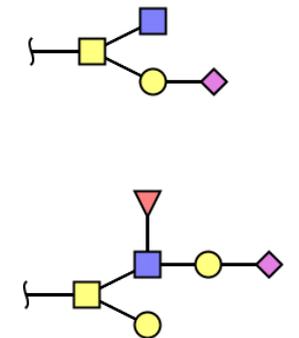
Sugar code:

- N-acetylglucosamine
- mannose
- galactose
- ◆ N-acetylneuraminic acid
- ◇ N-glycolylneuraminic acid
- ▲ fucose

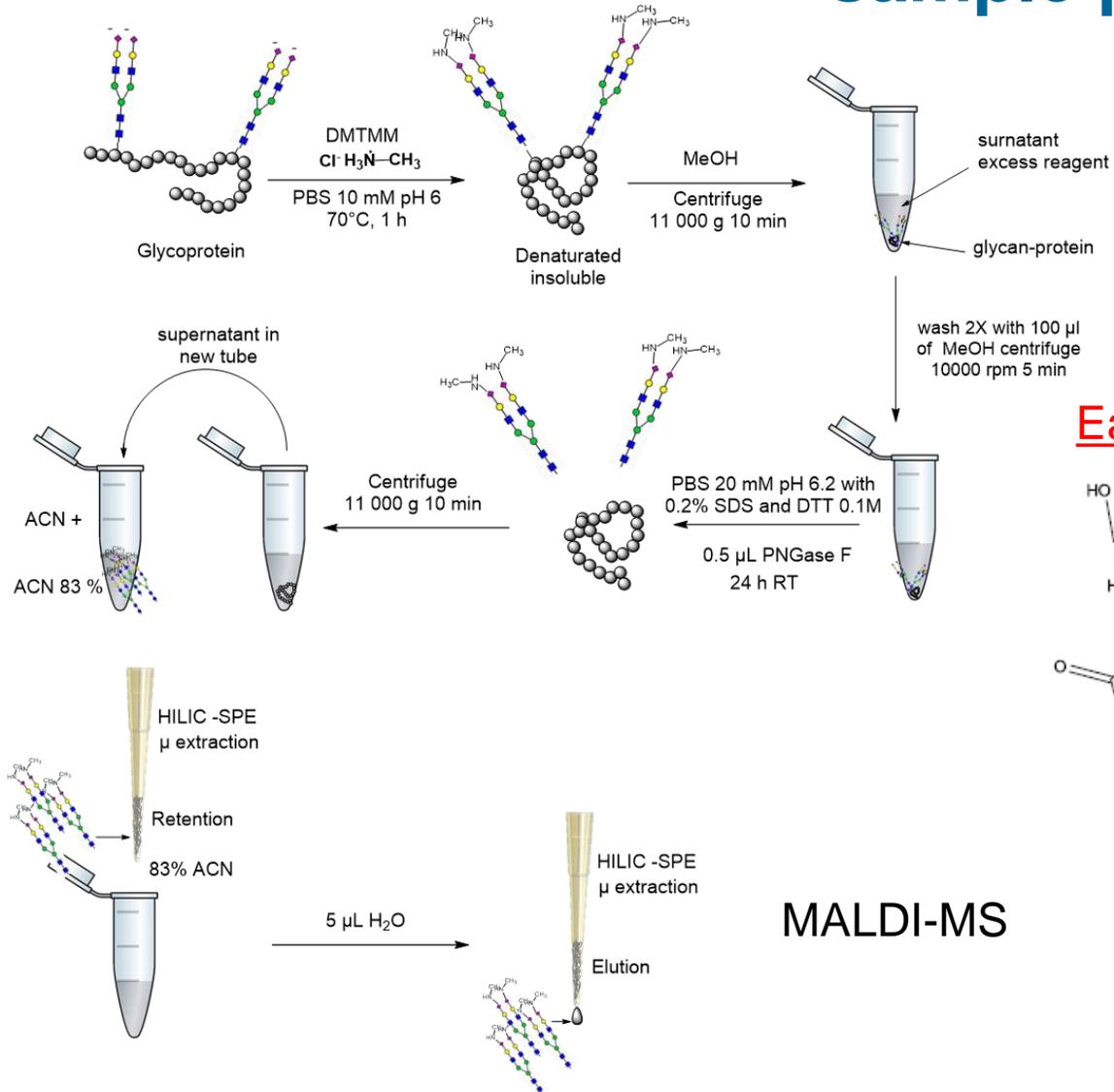
N-Glycan



O-Glycan

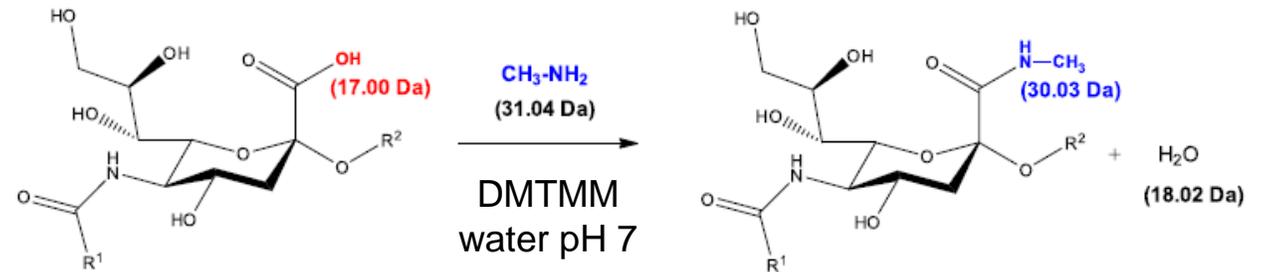


# N-Glycan analysis: sample preparation

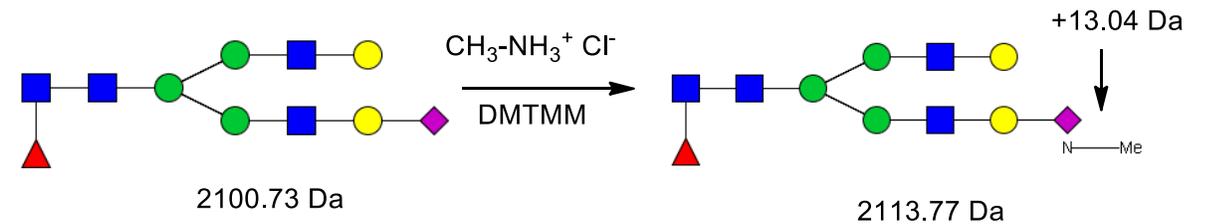


The stabilization of sialic acid by DMTMM-mediated methylamidation enables a conservation of the glycan structures, in contrast to other methods where the labile sialic acids are partially lost (e.g. methylation)

Each method highlight different aspects = difficult comparison

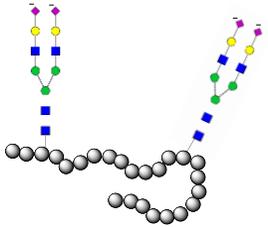


MALDI-MS

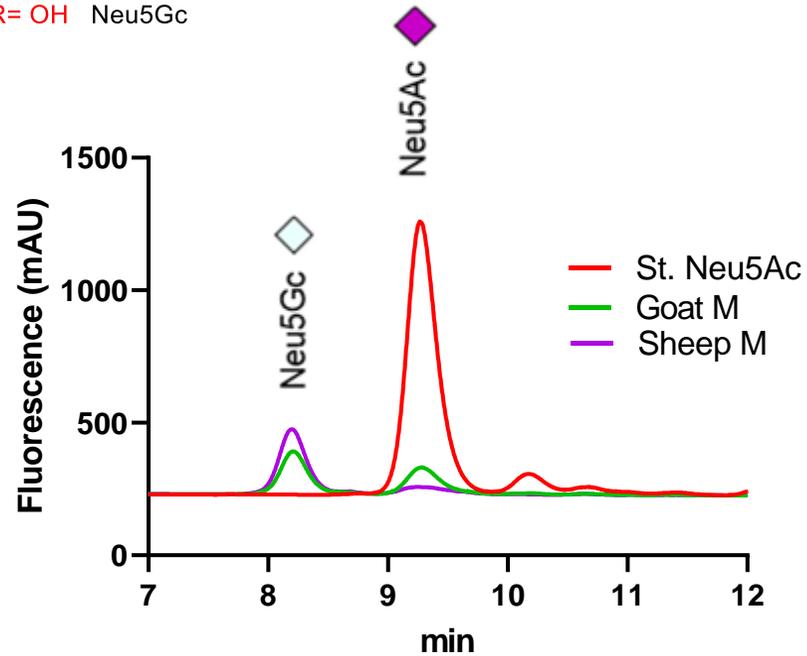
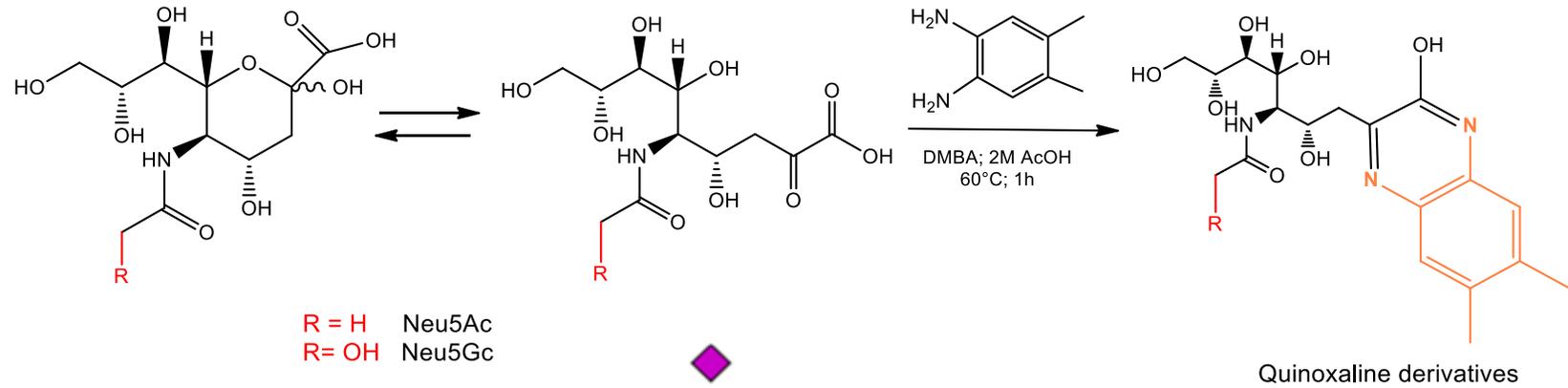


# Sialic acid quantification DMBA HPLC-FL detection

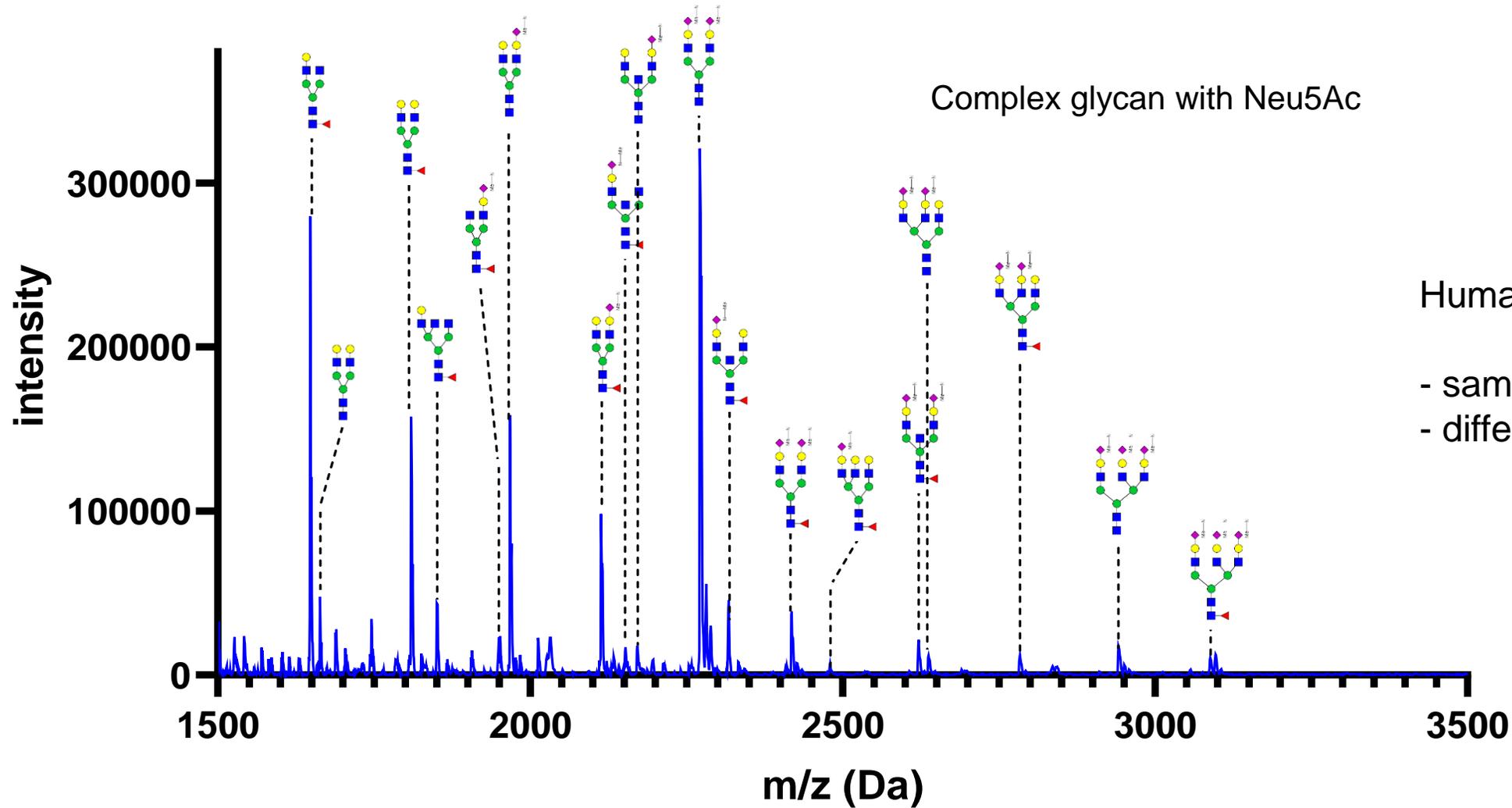
Glycoprotein



DMBA label  
HPLC-FL



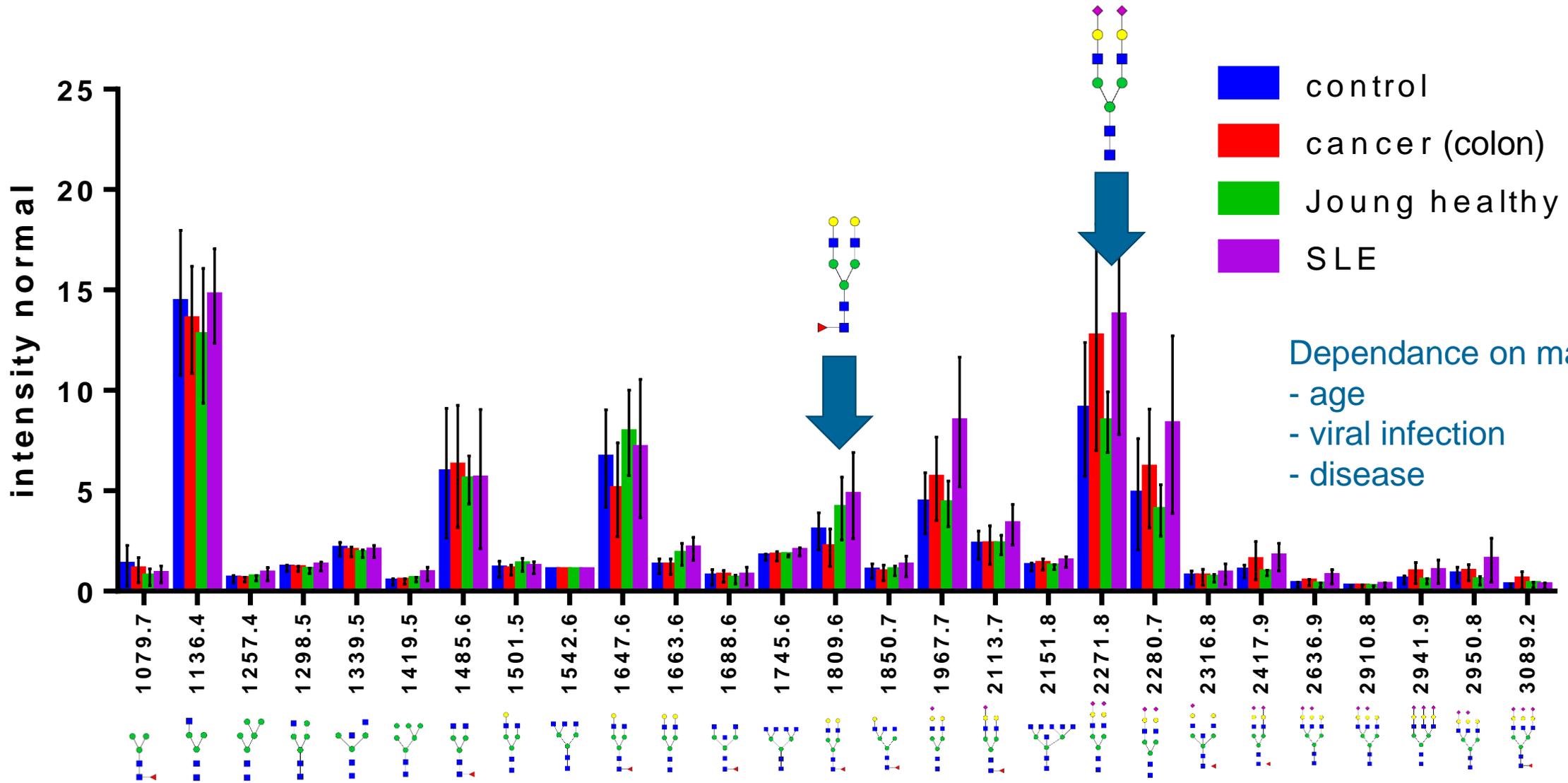
# Human total serum N-Glycans



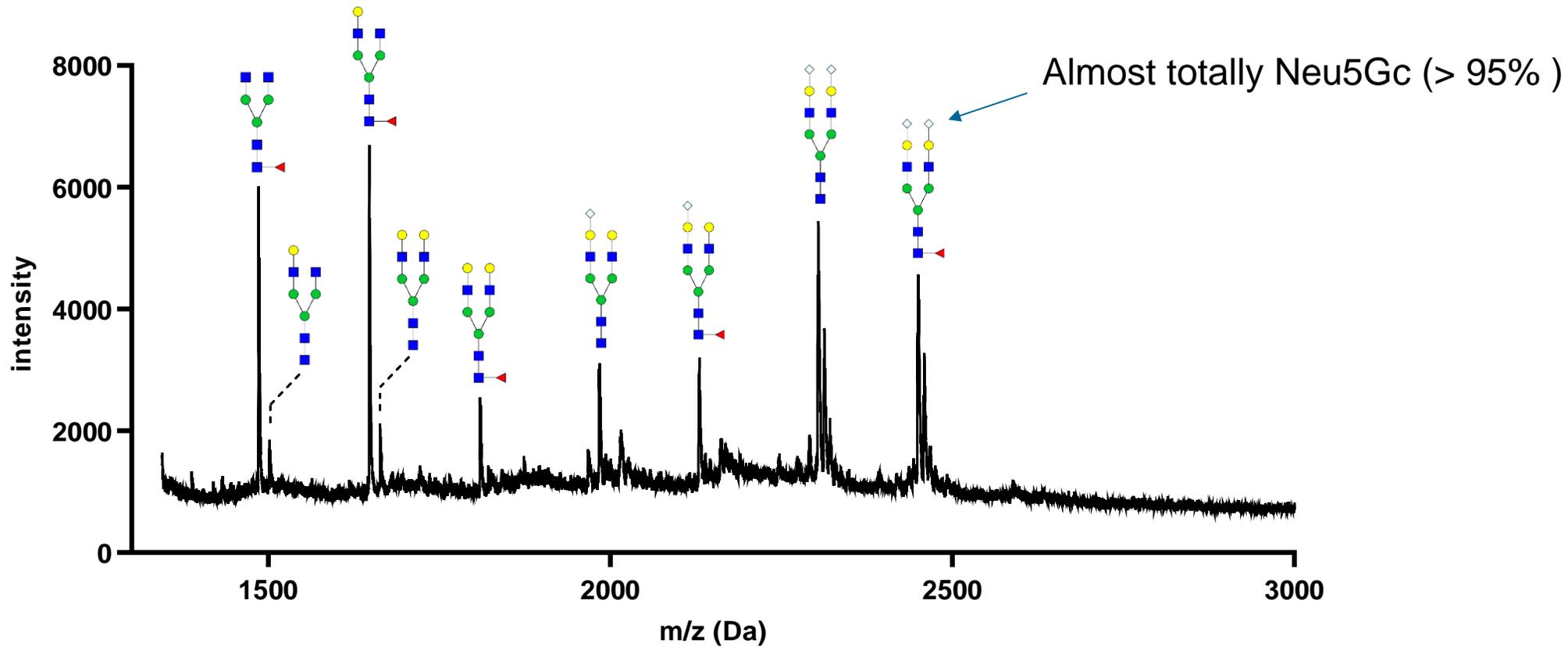
Human Serum N-glycans

- same glycans pattern
- different amount

# Serum N-glycan: Cancer, Systemic Lupus Erythematosus (SLE)



# Mouse total serum N-Glycans



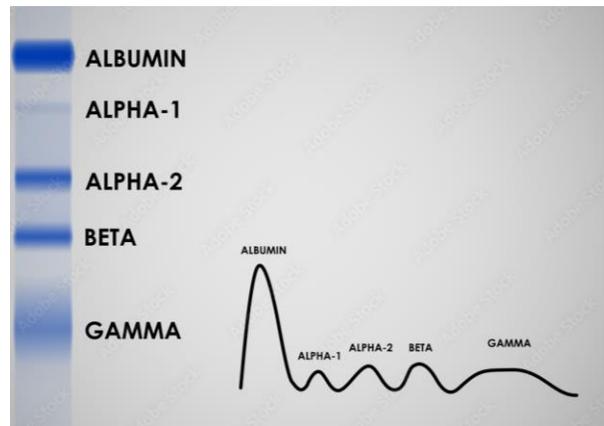
→ Mouse not good model for study human disease  
(not used for human virus study)

# N-glycan analysis

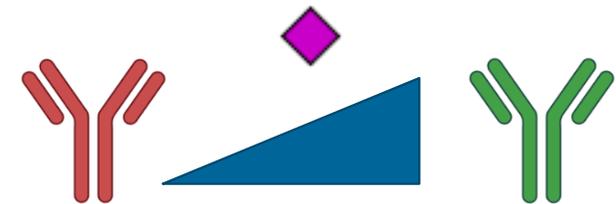
## From: total serum

We observed limited, but disease-specific variations between N-glycosylation of sera of patients with different pathologies.

## to tissue specific / fraction specific

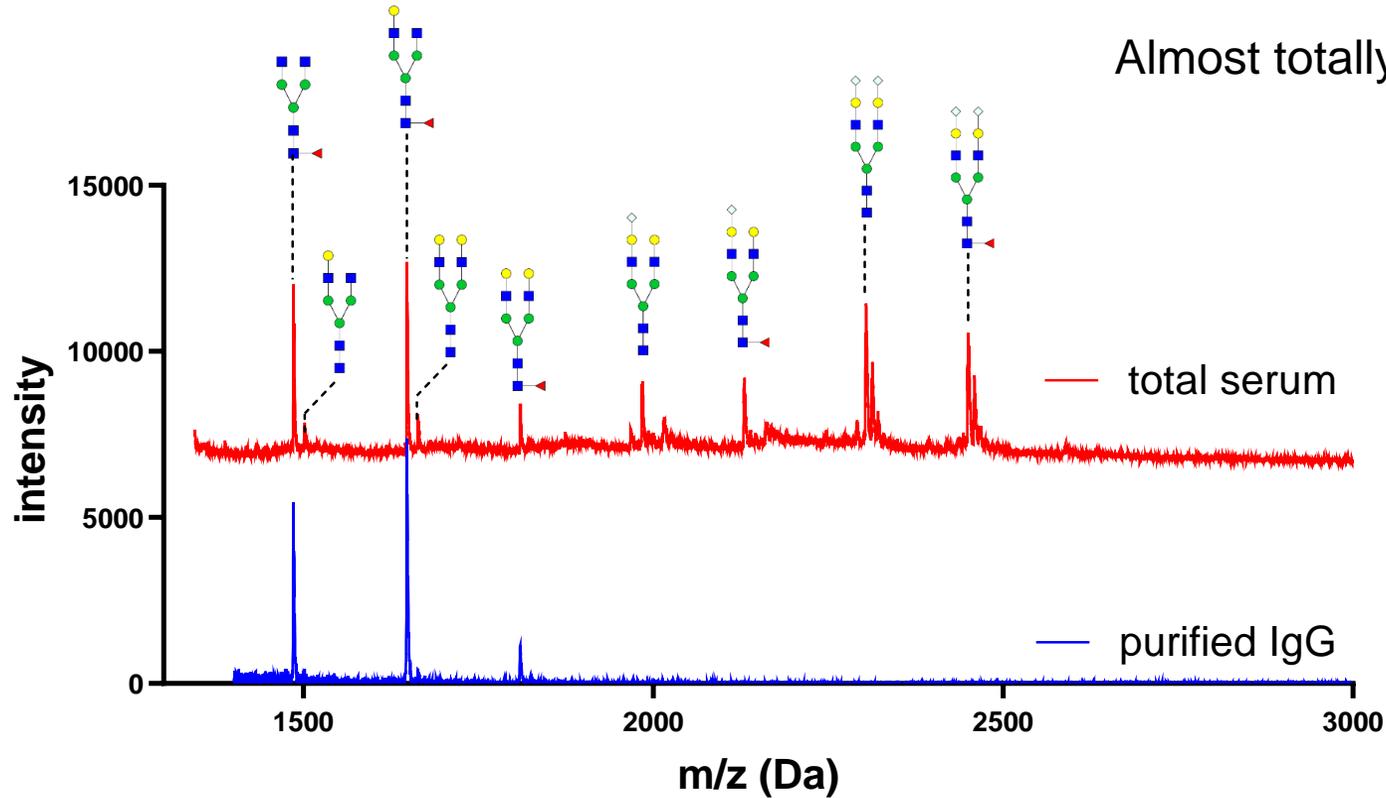


Pro inflammatory

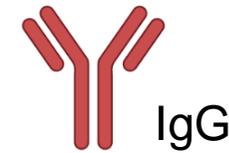


Anti inflammatory

# Mouse serum total and IgG fraction N-Glycans

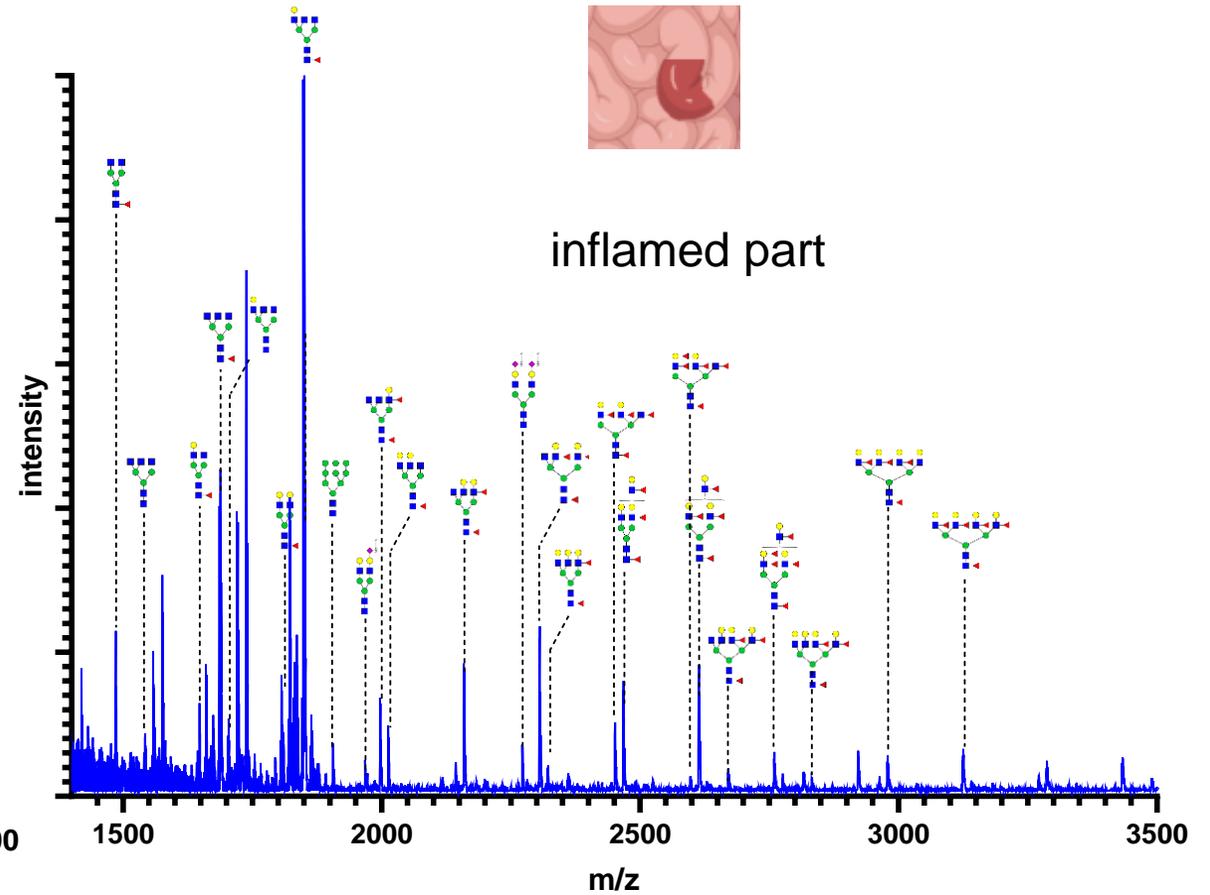
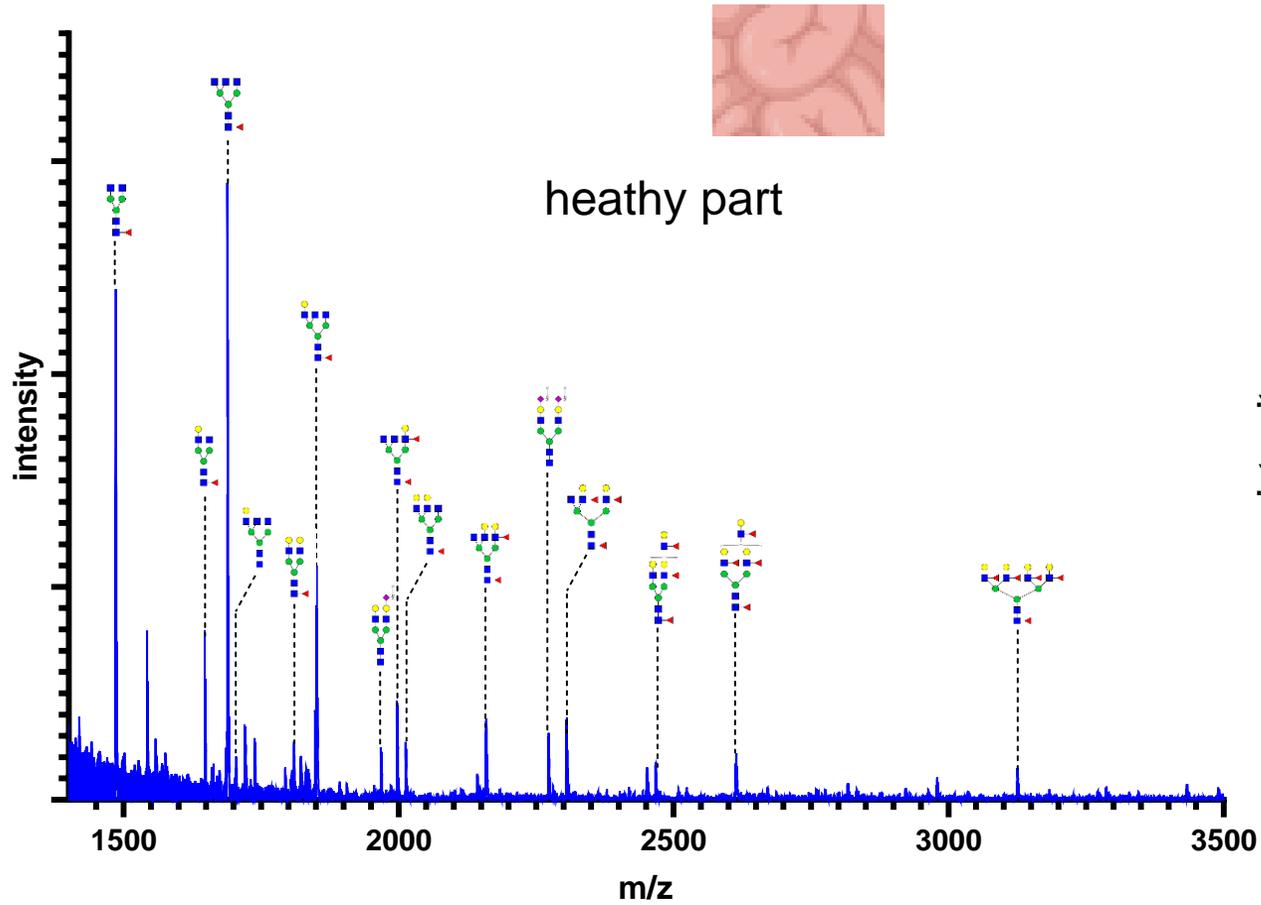


Almost totally Neu5Gc (> 95%)

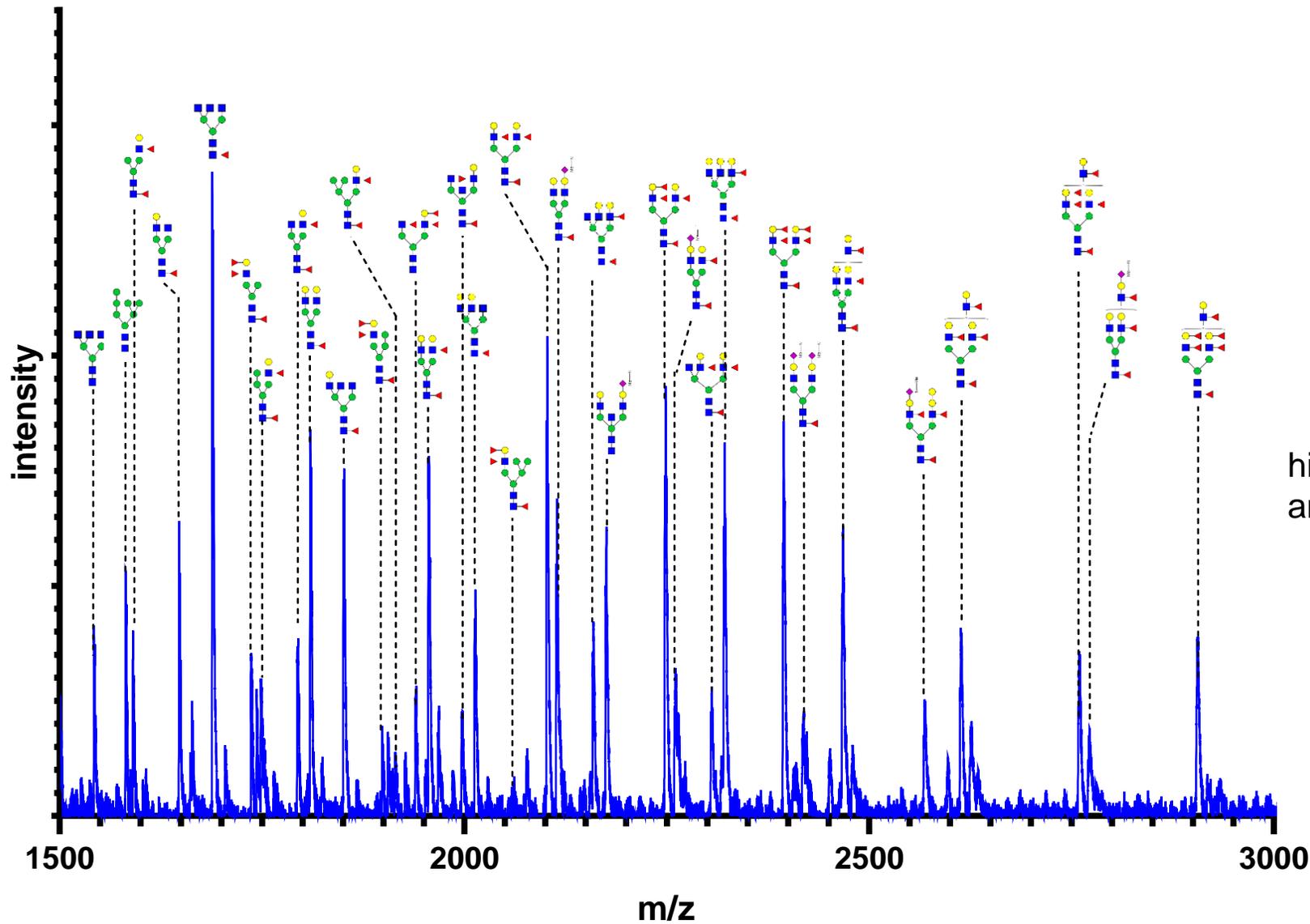
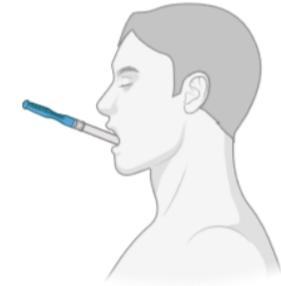


→ Mouse not good model for study human disease  
(e.g. not used for human virus study)

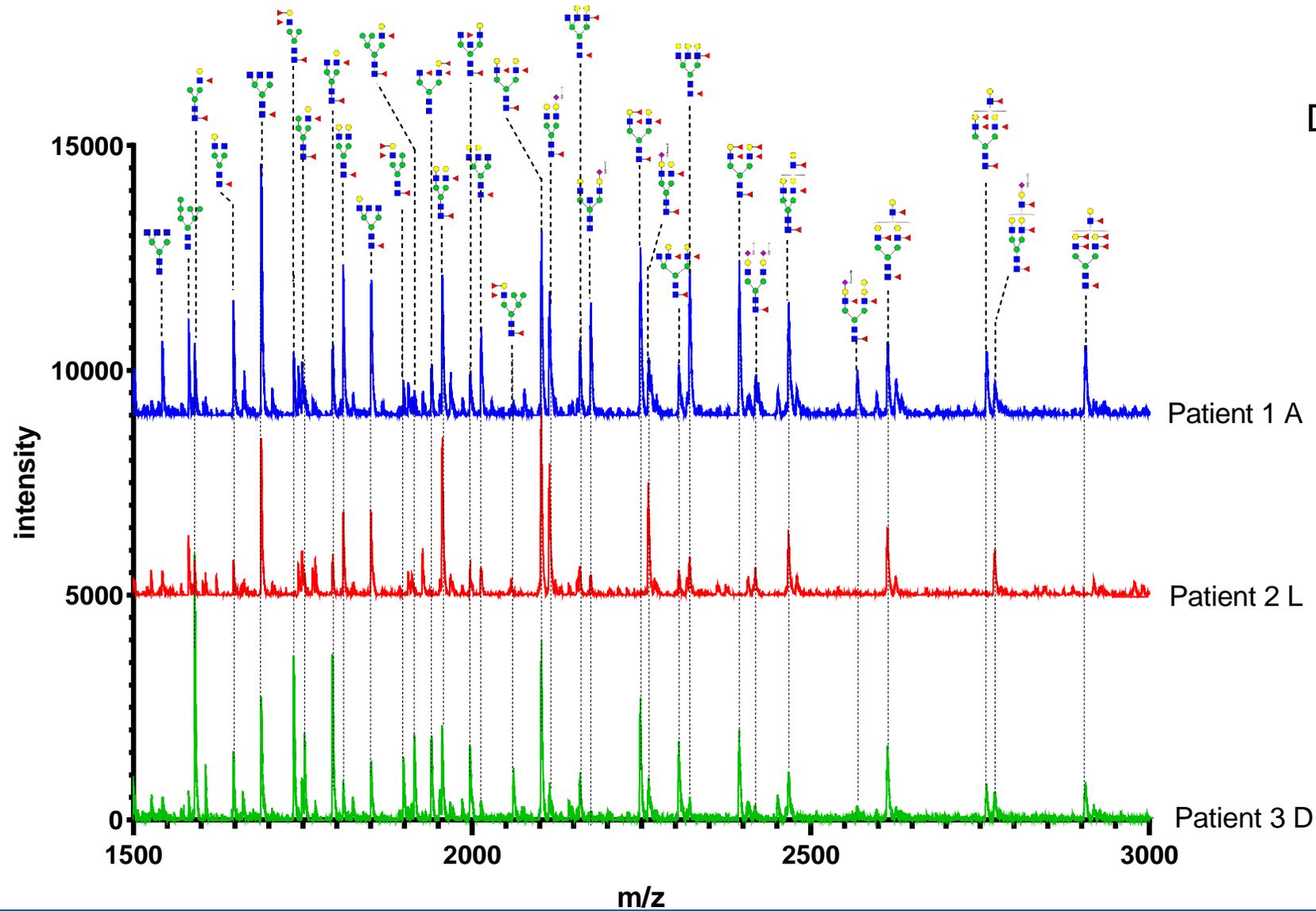
# Human intestine tissue N-glycans



# Human saliva N-glycans



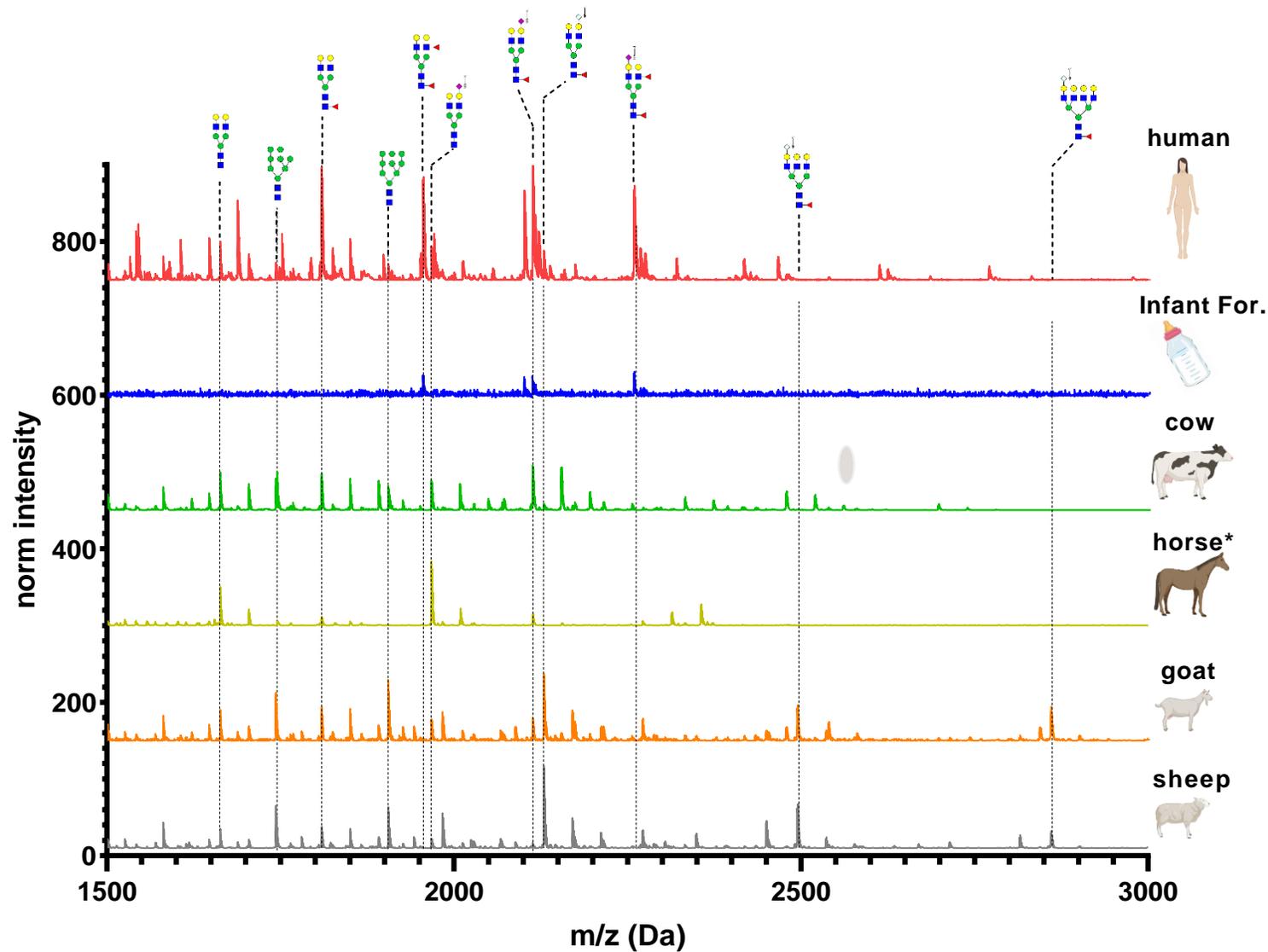
# Human saliva N-glycans



Different viral adhesion



# MALDI-MS spectra of milk of different species



Variability in the species



Variability between the species

# Conclusion

## total serum N-glycan analysis

**Species dependent** -> helpful for understand difference between animals / or animals model

We observed **limited, but disease-specific variations** between

N-glycosylation of sera of patients with different pathologies. (peak intensity)

## tissue specific / fraction specific

**biopsies, body fluid, antibody fraction, cell tissue** show a **greater N-glycan pattern diversity** between different patients and patients with different pathologies

-> best candidate for biomarker

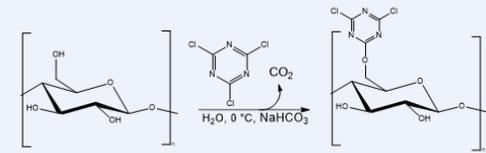
# Covalent protein immobilization on modified cellulose for N-glycan purification

Proteins are usually physically adsorbed onto supports, however they can spontaneously undergo desorption. By considering this, here is described a **covalent immobilization method**.



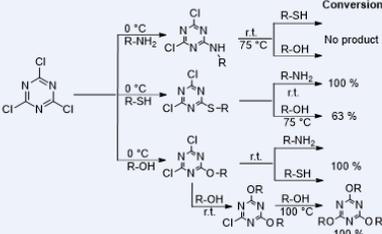
## Step 1: Modify

Cellulose functionalization with **trichloro triazine** for **covalent immobilization of proteins** and purification of **N-glycans**.



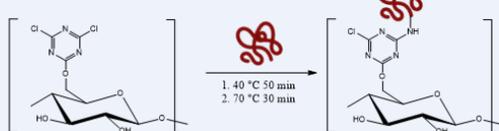
Substitution of chlorine atoms of trichloro triazine is possible at **different temperatures**.

Below: Temperature of reactions for subsequent substitution of chlorine atoms of trichloro triazine by different nucleophiles [2].

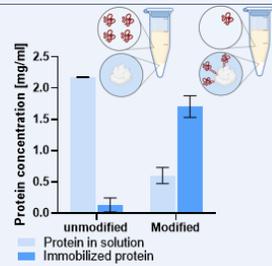


## Step 2: Immobilize

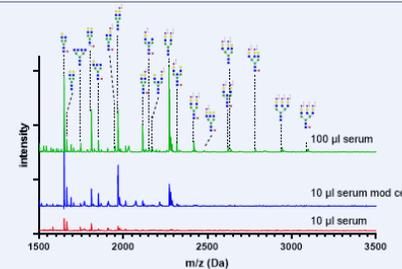
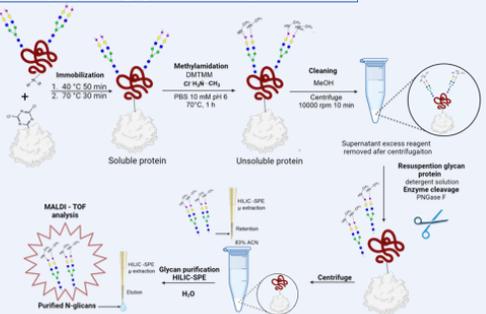
Protein immobilization via **covalent bond** between the **amine group** of the protein and the modified cellulose.



The degree of modification and the temperature of storage of cellulose are of key importance in the immobilization of the protein.



## Step 3: N-glycan analysis



By **immobilizing** the proteins, this method allows the analysis and quantification of glycans in **low amounts of proteins** (e.g. antibodies).



# Edible Bird's Nest: N-Glycosylation pattern and quantitative analysis of Sialic Acid

Lisa Ying<sup>a</sup>, Alessio Salvatore Gentile<sup>b</sup>, Daniel Garcia de Otazo Hernandez<sup>a</sup>, Haidi Jakovic<sup>a</sup>, Eva Untermayr<sup>b</sup>, Philipp Gritsch<sup>c</sup> and Davide Ret<sup>a</sup>

## BACKGROUND

In the Chinese community Edible Bird's Nest (EBN) has long been celebrated because of its nutritious properties. It contains a high amount of sialic acid among its various bioactive components. Sialic Acid is known for its nutritious values and has been widely researched. Its immune enhancing, anti-cancer as well as skin whitening properties are well acknowledged. For this reasons Sialic Acid is a promising functional ingredient in various fields. [1]



Figure 1: Edible Bird's Nest

Our aims are the quantification of total sialic acid amount in Edible Bird's Nest through HPLC-FL and the analysis of N-Glycosylation pattern with MALDI-ToF.

## N-GLYCOSYLATION PATTERN

1. Delipidation with Hexane and Isopropanol
2. Modification with methylamine using 4-(4,6-dimethoxy-1,3,5-triazin-2-yl)-4-methyl-morpholinium chloride (DMTMM) and methylamine hydrochloride
3. N-Glycan release by PNGase-F
4. Purification by HILIC solid phase extraction
5. Analysis of N-Glycans with MALDI-ToF/MS [2]



Figure 2: Reaction scheme of Methylation of sialic acid.

1. Cleavage of sialic acid from glycoconjugate using concentrated CH<sub>3</sub>COOH
2. Derivatization of sialic acid with 4,5-Dimethyl-1,2-Phenyldiamine (DMBA) and formation of fluorescent quinoxaline derivatives

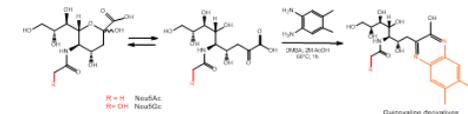


Figure 5: Reaction scheme of sialic acid with DMBA.

3. Analysis with HPLC-FL

## Results:

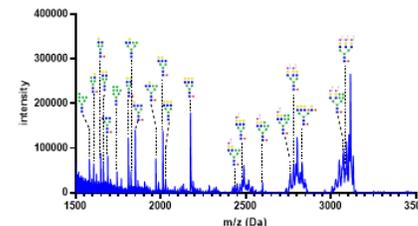


Figure 3: MALDI-ToF/MS spectra of the methylamidated N-glycan profile of EBN.

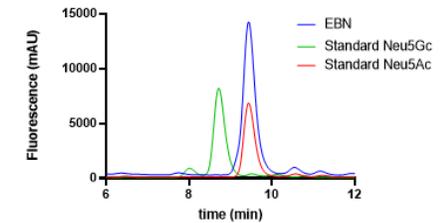


Figure 6: HPLC-Chromatograms of DMBA-tagged sialic acid. In EBN no Neu5Gc could be detected.

Table 1: The amount of Neu5Ac in EBN compared to cow milk.

sample	Neu5Ac	SD	unit
Cow milk	0.349	0.048	mg/mL
EBN	85.6	7.605	mg/g
One glass of cow milk (300 mL)	104.7	14.4	mg
One Leaf of EBN (6 g)	513.5	45.6	mg

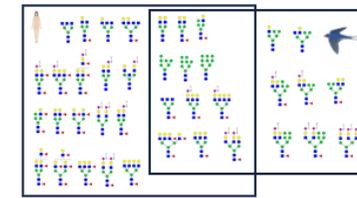


Figure 4: Venn plot of principal N-glycans in EBN compared to Human serum. High amount of hybrid type glycans are found in EBN.

## References:

- [1] Du, Y. et al. (2021) A comprehensive review of Edible Bird's Nest. *Food Research International*, 140, p. 109875.  
[2] Ret, D. et al. (2022) DMTMM-mediated methylation for MALDI mass spectrometry analysis of N-glycans with structurally conserved sialic residues. *Biological Fluids "Biofluids"*, Volume 242, p. 123326.

- a) Research Unit Macromolecular Chemistry, Institute of Applied Synthetic Chemistry, TU Wien  
b) Institute of Pathophysiology and Allergy Research, Center for Pathophysiology, Infectology and Immunology, Medical University of Vienna  
c) Research Unit Stereoselective and sustainable synthesis, Institute of Applied Synthetic Chemistry, TU Wien

# A robust quantification method of sialic acids and AGES precursors

Daniel García de Otazo Hernández<sup>a,b</sup>, Alessio Gentile<sup>b</sup>, Lisa Ying<sup>a</sup>, Haidi Jakovic<sup>a</sup>, Eva Untermayr<sup>a</sup>, Davide Retz<sup>a</sup>



Quinoxaline derivate stabilization allows for the reproducible quantification of diketone biomolecules present in raw and cooked foods.

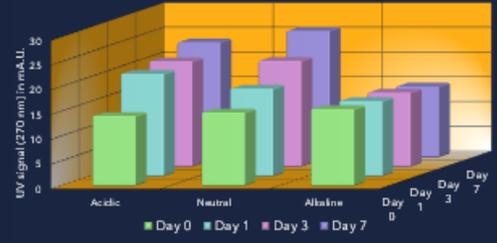
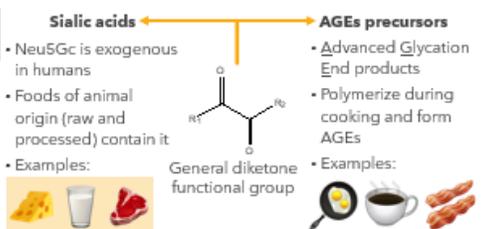


Fig.1: Neu5Ac + DMBA stability study (Quinoxaline derivate peak)

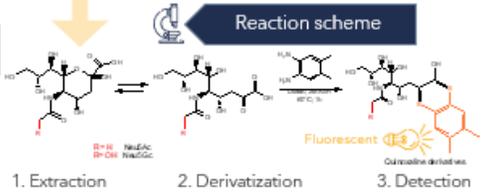
## What are diketone biomolecules?

Diketone biomolecules are substances containing two vicinal ketone functional groups. They are commonly found in neuraminic acids (Neu5Ac & Neu5Gc) as well as AGEs precursors (Glyoxal & Methylglyoxal). These diketone biomolecules are of special interest due to their negative impact on digestion in humans [1].



## How do we quantify them?

Starting from an organic matrix, sialic acids are first cleaved from the glycoprotein and then derivatized yielding a quinoxaline. AGEs precursors are extracted dependent on the sample matrix and derivatized the same way. The fluorescent quinoxaline-labelled analyte can now be easily quantified via HPLC-FL.



## Why is stabilizing necessary?

Previous analysis protocols suffered from a change in signal intensity and peak width when the quinoxaline derivate started to degrade over time. We have discovered that the derivate is stabilized by shifting to an alkaline pH after the reaction concludes. This way all possible side reactions are inhibited.

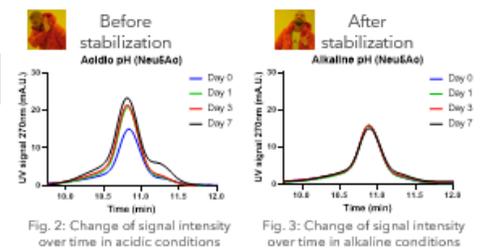
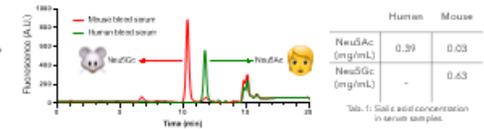


Fig. 2: Change of signal intensity over time in acidic conditions. Fig. 3: Change of signal intensity over time in alkaline conditions.

## Exemplary data:

Fig. 4: Measurement of sialic acids in human and mouse serum.



# Comparative N-Glycosylation Analysis in Human Serum: Insights into COVID-19, Long COVID, and ME/CFS

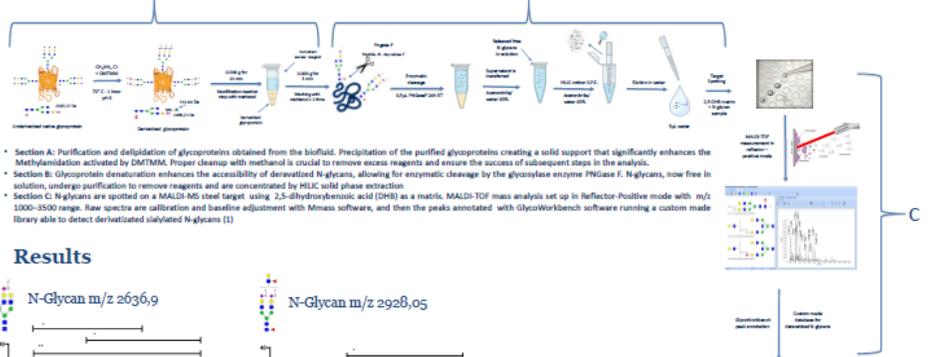
Davide Retz<sup>a,b</sup>, Salvatore Alessio Gentile<sup>b</sup>, Johanna Rohrhofer<sup>b</sup>, Larissa Koidl<sup>b</sup>, Daniel García de Otazo Hernández, Lisa Ying, Haidi Jakovic, Simone Knaus<sup>a</sup>, Eva Untermayr<sup>b</sup>

<sup>a</sup> Research Unit Macromolecular Chemistry, Institute of Applied Synthetic Chemistry, TU Wien, 1060 Vienna, Austria, <sup>b</sup> Institute of Pathophysiology and Allergy Research, Center for Pathophysiology, Infectiology and Immunology, Medical University of Vienna, 1090 Vienna, Austria

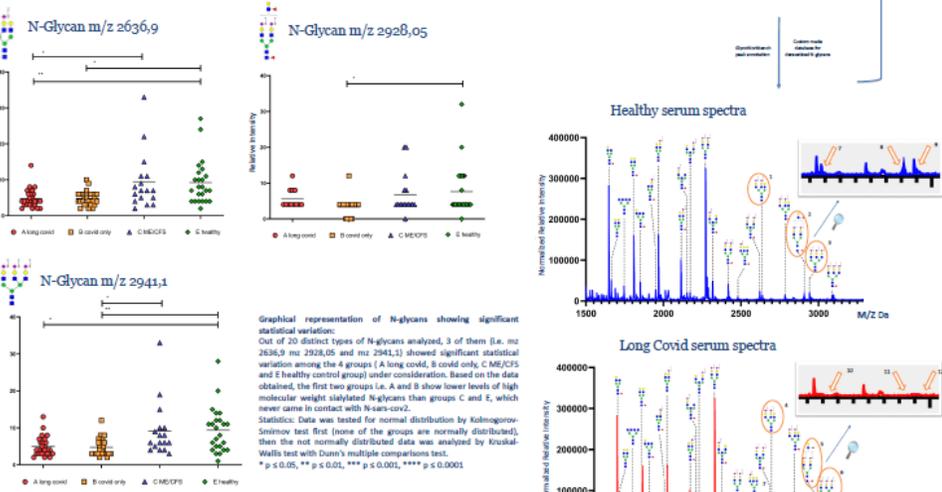
## Why N-Glycosylation?

Evaluation of N-glycosylation pattern is gaining interest in immunology as it plays a crucial role in the structure and function of molecules relevant for the immune system. N-glycans can modulate the activity and specificity of antibodies, the complement system as well as pathogen-host interactions influencing the ability of antigens recognition and of eliciting an immune response. Alterations in N-glycan structures have implications in a large variety of diseases such as rheumatoid arthritis, cancers and viral infection. Thus, a comprehensive picture of the N-glycosylation profile is pivotal for better understanding of mechanisms associated with poorly understood disorders such as post-viral fatigue.

## Method



## Results



## Findings

- Our method allows a rapid, accurate and cost-effective identification of N-glycans in biofluids opening up future developments in diagnostics. New biomarkers can be identified, since distinct N-glycosylation patterns have been observed in many pathophysiological conditions.
- In patients' sera more than 10 weeks after COVID infection we observed significant changes in the N-glycome profile. High molecular weight glycans were reduced significantly in patients after COVID infections being associated with elevated levels of non-sialylated glycans. This was in contrast to the pattern detected in healthy controls indicating a persistent change of the glycome on serum proteins after viral infection.
- Even though the ME/CFS group C presented chronic fatigue symptoms similar to post covid fatigue and had developed disease also after an infection (2), the N-glycosylation patterns were similar to those of the healthy control group. This indicated that N-glycosylation pattern restore over time.

**References**  
 1) Retz D. et al, *Talanta*, May 2022  
 2) Rohrhofer J. et al, *Allergy* 2023



**TEAM MEMBERS TU Wien:**

**Daniel García de Otazo Hernández  
Lisa Ying  
Haidi Jakovic  
Antonio Ammaturo  
Veronica Monti  
Sabatino Gianluigi  
Philipp Gritsch  
Knaus Simone**

**TEAM MEMBERS Meduni Wien:**

**Gentile Alessio  
Johanna Rohrhofer  
Larissa Koidl  
Untersmayr Eva**



**Collaborators Meduni Graz:**

**Selina Keppler  
Matteo Villa**

**Collaborators Meduni Wien:**

**Arvand Haschemi**



REGIONE AUTONOMA  
FRIULI VENEZIA GIULIA

**Stefania Garofalo**

**Thank you for your attention**